Advances in Experimental Medicine and Biology 1326 Cell Biology and Translational Medicine

Kursad Turksen Editor

Cell Biology and Translational Medicine, Volume 12

Stem Cells in Development and Disease



Advances in Experimental Medicine and Biology

Cell Biology and Translational Medicine

Volume 1326

Series Editor

Kursad Turksen (emeritus), Ottawa Hospital Research Institute, Ottawa, ON, Canada

Editorial Board

Pascal Pineau, Institut Pasteur, Paris, France Daisuke Sugiyama, Kyushu University, Fukuoka, Japan Jeffrey M. Gimble, Louisiana State University, Baton Rouge, LA, USA Pablo Menendez, Josep Carreras Leukaemia Research Institut, Barcelona, Spain Cesar V. Borlongan, University of South Florida Health, Tampa, FL, USA Essam M. Abdelalim, Diabetes Research Institute, Doha, Qatar Aaron W. James, Johns Hopkins Hospital, Baltimore, MD, USA Srikala Raghavan, Institute for Stem Cell Science and Regenerative Medicine, Bengaluru, Karnataka, India Tiziana A. L. Brevini, University of Milan, Milan, Italy Murat Y. Elcin, Ankara University, Ankara, Turkey Mario Tiberi, Ottawa Hospital, Ottawa, ON, Canada Nagwa El-Badri, Zewail City of Science and Technology, Giza, Egypt Panos Kouklis, University of Ioannina, Mpizani, Greece Benjamin Levi, The University of Texas Southwestern Medical Center, Dallas, TX, USA

Cell Biology and Translational Medicine aims to publish articles that integrate the current advances in Cell Biology research with the latest developments in Translational Medicine. It is the latest subseries in the highly successful Advances in Experimental Medicine and Biology book series and provides a publication vehicle for articles focusing on new developments, methods and research, as well as opinions and principles. The Series will cover both basic and applied research of the cell and its organelles' structural and functional roles, physiology, signalling, cell stress, cell-cell communications, and its applications to the diagnosis and therapy of disease.

Individual volumes may include topics covering any aspect of life sciences and biomedicine e.g. cell biology, translational medicine, stem cell research, biochemistry, biophysics, regenerative medicine, immunology, molecular biology, and genetics. However, manuscripts will be selected on the basis of their contribution and advancement of our understanding of cell biology and its advancement in translational medicine. Each volume will focus on a specific topic as selected by the Editor. All submitted manuscripts shall be reviewed by the Editor provided they are related to the theme of the volume. Accepted articles will be published online no later than two months following acceptance.

More information about this subseries at http://www.springer.com/series/15838

Kursad Turksen Editor

Cell Biology and Translational Medicine, Volume 12

Stem Cells in Development and Disease



Editor Kursad Turksen *(emeritus)* Ottawa Hospital Research Institute Ottawa, ON, Canada

ISSN 0065-2598 ISSN 2214-8019 (electronic) Advances in Experimental Medicine and Biology ISSN 2522-090X ISSN 2522-0918 (electronic) Cell Biology and Translational Medicine ISBN 978-3-030-71932-6 ISBN 978-3-030-71933-3 (eBook) https://doi.org/10.1007/978-3-030-71933-3

 ${\rm (}{\rm \bigcirc}$ The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Switzerland AG 2021

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed. The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use. The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication.

this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Switzerland AG The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland

Preface

This next volume in the Cell Biology and Translational Medicine series addresses the topic of stem cells in development and diseases. Amongst specialized topics, there are chapters on the role of stem cells neuronal development and the role of stem cells in diseases including arthritis, aging and cancer.

I remain very grateful to Gonzalo Cordova, associate editor of the series, and acknowledge his continuous support.

I would also like to acknowledge and thank Mariska van der Stigchel, assistant editor, for her outstanding efforts in helping to get this volume to the production stages.

A special thank you goes to Shanthi Ramamoorthy and Rathika Ramkumar for their outstanding efforts in the production of this volume.

Finally, sincere thanks to the contributors not only for their support of the series, but also for their insights and efforts to capture both the advances and the remaining obstacles in their areas of research. I trust readers will find their contributions as interesting and helpful as I have.

Ottawa, ON, Canada

Kursad Turksen

Contents

Immune Dysregulation and Recurring Mutationsin Myelodysplastic Syndromes PathogenesisAnacélia Matos, Silvia M. M. Magalhães, and Michael J. Rauh	1
Stem Cell Aging and Regenerative Medicine Debojyoti De, Parimal Karmakar, and Debalina Bhattacharya	11
Potential of Chimeric Antigen Receptor T-Cells in Cancer Therapy	39
Drug Sensitivity and Drug Repurposing Platform for Cancer Precision Medicine	47
Role and Regulation of Lin28 in ProgenitorCells During Central Nervous System Development	55
Cartilage Repair by Mesenchymal Stem Cell-Derived Exosomes: Preclinical and Clinical Trial Update and Perspectives	73
Kaempferol Induces Cell Death and Sensitizes HumanHead and Neck Squamous Cell Carcinoma CellLines to CisplatinMabel Catalán, Catalina Rodríguez, Ivonne Olmedo,Javiera Carrasco-Rojas, Diego Rojas, Alfredo Molina-Berríos,Mario Díaz-Dosque, and José A. Jara	95
A Comprehensive Approach to Urticaria: From ClinicalPresentation to Modern Biological Treatments ThroughPathogenesisPathogenesis1Marco Folci, Giacomo Ramponi, and Enrico Brunetta	11

The Potentials and Pitfalls of Using Adult Stem Cells			
in Cancer Treatment			
Mrinal K. Das, Taral R. Lunavat, Hrvoje Miletic,			
and Jubayer A. Hossain			
The Outcome of Stem Cell-Based Therapies on the Immune			
Responses in Rheumatoid Arthritis	159		
Peyvand Parhizkar Roudsari, Sepideh Alavi-Moghadam,			
Mostafa Rezaei-Tavirani, Parisa Goodarzi, Akram Tayanloo-Beik,			
Forough Azam Sayahpour, Bagher Larijani, and Babak Arjmand			
Index	187		



Immune Dysregulation and Recurring Mutations in Myelodysplastic Syndromes Pathogenesis

Anacélia Matos, Silvia M. M. Magalhães, and Michael J. Rauh

Abstract

Myelodysplastic syndromes (MDS) are clonal stem cell malignancies characterized by ineffective hematopoiesis leading to peripheral cytopenias and variable risk of progression to acute myeloid leukemia. Inflammation is associated with MDS pathogenesis. Several cytokines, reactive species of oxygen/nitrogen and growth factors are directly or indirectly involved in dysfunction of the MDS bone marrow (BM) microenvironment. Mutations in genes mainly regulating RNA splicing, DNA methylation and chromatin accessibility, transcription factors, signal transduction and

Authors Silvia M. M. Magalhães and Michael J. Rauh have equally contributed to this chapter.

A. Matos

Cancer Cytogenomic Laboratory, Federal University of Ceará, Fortaleza, Brazil

Post-graduate Program in Medical Science, Federal University of Ceará, Fortaleza, Brazil

S. M. M. Magalhães Cancer Cytogenomic Laboratory, Federal University of Ceará, Fortaleza, Brazil

Post-graduate Program in Medical Science, Federal University of Ceará, Fortaleza, Brazil

Department of Clinical Medicine, Federal University of Ceará, Fortaleza, Brazil e-mail: silviamm@ufc.br

M. J. Rauh (🖂)

Department of Pathology and Molecular Medicine, Queen's University, Kingston, ON, Canada e-mail: michael.rauh@queensu.ca; rauhm@queensu.ca the response to DNA damage contribute to ineffective hematopoiesis, genomic instability and MDS development. The inflammationassociated DNA damage in hematopoietic stem cells may also contribute to MDS development and progression with aggressive clinical characteristics. Many studies have aimed at clarifying mechanisms involved in the activity of immature myeloid cells as powerful modulators of the immune response and their correlation with aging, autoimmunity, and development of cancer. In this review, we explore recent advances and accumulating evidence uniting immune dysregulation, inflammaging and recurring mutations in the pathogenesis of MDS.

Keywords

Bone marrow \cdot Hematopoietic stem cells \cdot Inflammation \cdot Mutations \cdot Myelodysplastic syndromes

Abbreviations

AML	acute myeloid leukemia
ARG1	arginase 1
ASXL1	Additional Sex Combs Like 1, Tran-
	scriptional Regulator
BM	bone marrow

CBL	Casitas B-Lineage Lymphoma	RAD21	RAD21 Cohesin Complex	
	Proto-Oncogene		Component	
CCUS	clonal cytopenia of undetermined	RNA	ribonucleic acid	
	significance	ROS	reactive oxygen species	
CD	cluster of differentiation	RUNX1	RUNX Family Transcription	
CHIP	clonal hematopoiesis of indetermi-		Factor 1	
	nate potential	S100A8	S100 Calcium Binding Protein A8	
CSF	colony-stimulating factor	S100A9	S100 Calcium Binding Protein A9	
DAMP	danger-associated molecular pattern	SF3B1	Splicing Factor 3b Subunit 1	
del	deletion	SRSF2	Serine And Arginine Rich Splicing	
DNA	deoxyribonucleic acid		Factor 2	
DNMT3A	DNA methyltransferase 3A	STAG2	Stromal Antigen 2	
eMDSC	early MDSC	TAM	tumour-associated macrophage	
ETV6	ETS Variant Transcription Factor 6	TCR	T-cell receptor	
EZH2	Enhancer Of Zeste 2 Polycomb	TET2	Ten-Eleven Translocation	
	Repressive Complex 2 Subunit		Methylcytosine Dioxygenase 2	
G	granulocytic	TGF-β	transforming growth factor beta	
GATA2	GATA Binding Protein 2	TIFAB	TRAF-Interacting Protein with	
GM	granulocyte-monocyte		Forkhead-Associated Domain,	
HLA	human leukocyte antigen		Family Member B	
HSPC	hematopoietic stem/progenitor cell	TIRAP	Toll-interleukin-1 receptor domain-	
IDH1/2	isocitrate dehydrogenases 1 and 2		containing adaptor protein	
IFN-γ	interferon gamma	TLR	Toll-like receptor	
IL .	interleukin	TME	tumour microenvironment	
IMC	immature myeloid cell	TNF-α	tumour necrosis factor alpha	
iNOS	inducible nitric oxide synthase	TP53	Tumour Protein P53	
IPSS	International prognostic scoring	TRAF	tumor necrosis factor receptor-	
	system		associated factor	
JAK2	Janus Kinase 2	Treg	regulatory T-cell	
L-Arg	L-arginine	U2AF1	U2 Small Nuclear RNA Auxiliary	
М	monocytic		Factor 1	
MDS	myelodysplastic syndromes	VEGF	vascular endothelial growth factor	
MDSC	myeloid-derived suppressor cell	WT1	Wilms tumour 1	
miR	micro RNA	ZRSR2	Zinc Finger CCCH-Type, RNA	
NF-κB	nuclear factor kappa-light-chain-		Binding Motif And Serine/Arginine	
	enhancer of activated B cells		Rich 2	
NK	natural killer			
NKT	natural killer/T			
NO	nitric oxide	1 In	troduction: Immune	
NRAS	Neuroblastoma RAS Viral (V-Ras)	D	vsregulation in MDS	
	Oncogene Homolog			
PAMP	pathogen-associated molecular	Myelodysp	lastic syndromes (MDS) constitute a	
-	pattern	group of di	seases which are distinguished by the	
PGE2	prostaglandin E2	presence of	f one or more unexplained peripheral	
PMN	polymorphonuclear	blood cytor	penias, dysplastic hematopoietic dif-	
PPM1D	Protein Phosphatase. Mg2+/Mn2+	ferentiation, and variable risk to progress to		
	Dependent 1D	acute myel	oid leukemia (AML). The incidence	

of these disorders increases with age, with an average of 70 years (Sekeres 2010). Recurrent mutations and haploinsufficiency of particular genes, related epigenetic changes, altered RNA splicing, and disorder in the bone marrow micro-environment all contribute to the disease phenotype (Cazzola 2020).

Inflammation is involved in many disease processes, such as hypertension, cardiovascular disease, rheumatoid arthritis, rheumatoid heart disease, and systemic lupus erythematosus, which are characterized by impairment of immune cell regulatory mechanisms. Several studies have focused on understanding the immunological abnormalities in in MDS (Banerjee et al. 2019; Xin et al. 2019; Corey et al. 2007; Rosenberg and Sinha 2009; Yang et al. 2015).

The pathogenesis of MDS is heterogeneous and includes abnormalities of both innate and adaptive immune systems, as will be described. Understanding how senescence-dependent changes and mutations impact both hematopoietic stem/progenitor cells (HSPC) and the bone marrow microenvironment is essential to understanding the pathogenesis and progression of the disease (Xin et al. 2019; Wang et al. 2018; Glenthoj et al. 2016; Kornblau et al. 2010; Marvel and Gabrilovich 2015).

The innate immune system was traditionally though to dysregulate HSPC proliferation and trigger apoptotic events contributing to the hallmark ineffective hematopoiesis in MDS. The activation of innate immune cells happens through the interaction between pathogen-associated molecular patterns (PAMPs) or host cell-derived danger-associated molecular patterns (DAMPs) with the Toll-like receptors (TLRs) (Kawai and Akira 2010). The TLR signaling pathway results in activation of mitogen-activated protein kinase cascades and the nuclear factor kappa-lightchain-enhancer of activated B cells (NF-kB) and leads to transcription of pro-inflammatory cytokines such as interleukin-8, which has been described by de Matos et al. (2017). Intrinsic dysregulation of TLR pathways in MDS HSPC results in hyperactive TLR signalling, including a novel inflammatory form of programmed cell death, known as pyroptosis (Basiorka et al.

2016; Barreyro et al. 2018). Moreover, MDS HSPC have a competitive advantage over normal HSPC in the resultant chronic inflammatory environment, which favours their expansion and disease progression (Muto et al. 2020).

Regarding cytokines, the levels of TNF- α , IFN- γ , TGF- β , IL-6 and IL-8 have been observed to be higher in MDS patients, and associated with both dysregulated inflammatory signaling and myeloid differentiation (Yang et al. 2015; Wang et al. 2018; Kornblau et al. 2010; de Matos et al. 2017). According to Xin et al. (2019), who conducted the first meta-analysis focused on inflammatory cytokine levels in MDS patients, there is a close association between immunological microenvironment disorders and the pathogenesis of MDS, with significantly increased TNF- α , IFN- γ , IL-6 and IL-8 in blood and bone marrow of MDS patients (Xin et al. 2019). In intrinsic addition to the aforementioned dysregulation of inflammatory pathways in HPSC, extrinsic influence of these cytokines from chronic infections or host inflammatory disorders may further foster the MDS clonal advantage and disease progression.

MDS-associated mutations and cytogenetic aberrations may also contribute to the inflammatory milieu. In a study made by Kornblau et al. (2010), some cytokines and chemokines were strongly correlated with certain MDS and AML cytogenetic abnormalities, and influenced MDS outcomes beyond the IPSS-calculated risk (Kornblau et al. 2010). The groups of Karsan and Starczynowski have also demonstrated that haploinsufficiency of micro-RNAs (e.g. miR-145 and miR-146a) and genes in del(5q) MDS (e.g. TIFAB) contribute to inappropriate TLR activation, including IL-6 production, by affecting expression of signalling mediators, Toll-interleukin-1 receptor domain-containing adaptor protein (TIRAP) and tumor necrosis factor receptorassociated factor-6 (TRAF6) (Starczynowski et al. 2010; Varney et al. 2015). Moreover, recurrent MDS-associated mutations in epigenetic regulators (e.g. TET2, ASXL1) and components of the spliceosome machinery (e.g. SF3B1, SRSF2, U2AF1) also appear to converge on innate immune pathways, resulting in excessive inflammasome activation and inflammatory cytokine production, including IL-6 (Basiorka et al. 2016; Smith et al. 2019; Pollyea et al. 2019).

The expression of at least thirty cytokines, chemokines and growth factors in the peripheral blood and bone marrow have been implicated in MDS pathogenesis and clinical outcomes (Banerjee et al. 2019; Xin et al. 2019; Yang et al. 2015; Wang et al. 2018; Glenthoj et al. 2016; Kornblau et al. 2010; Ganan-Gomez et al. 2015). Increased levels of cell death in bone marrow are a hallmark of lower risk disease. On the contrary, in higher risk MDS, with more aggressive clonal expansion, decreased levels of apoptosis are observed (Yang et al. 2015; Glenthoj et al. 2016; Kerbauy and Joachim 2007). Levels of IFN- γ and IL-6 are associated with apoptosis induction in the bone marrow of MDS patients and higher IFN-y and IL-6 secretion is generally related to lower-risk MDS. In contrast, immunosuppressive cytokines like IL-10 are more strongly secreted in high-risk MDS (Wang et al. 2018). These studies reflect the importance of inflammatory cytokines in dysregulation of the immunological environment in the pathogenesis of MDS. While they show a range in cytokine profiles between different types of MDS and different studies, they importantly also demonstrate convergence in critical innate immune pathways. Increased intramedullary apoptosis and pyroptosis are important contributors to cytopenias in MDS. Additionally, it is very probable that these immunologic aberrations and pressures play a central role in the course of MDS evolution from low- to high-risk MDS or to AML (Banerjee et al. 2019; Glenthoj et al. 2016; Steensma et al. 2015; Valka et al. 2019; Sallman and List 2019).

The role of adaptive immunity in MDS pathogenesis and progression has been considered, although not as extensively as the molecular connections with innate immunity. CD8+ T-cells may become activated and expanded in response to epitopes on MDS stem cells, resulting in suppression of both malignant and normal hematopoiesis, as reviewed by Wang et al. (2018). This is exemplified in lower-risk MDS with trisomy 8, where the Wilms tumor 1 antigen (WT1) is overexpressed by HPSC, WT1 triggers T-cell suppression of hematopoiesis, and this may be ameliorated by T-cell directed immunosuppressive therapy (Sloand et al. 2011). However, as MDS progresses to high-risk and later stages, there is expansion of regulatory T-cell subsets (Treg) and increased expression of inhibitory checkpoint proteins, which likely facilitate evasion of adaptive immunity by mutant MDS clones (see recent reviews (Wang et al. 2018; Barreyro et al. 2018; Sallman and List 2019; Winter et al. 2020)).

Indeed, it is well described that the tumor microenvironment in MDS and other cancers is immunosuppressive, both inhibiting activated immune cells and activating cells with a suppressive phenotype. Multiple cell types contribute to tumor mediated immune suppression, including Treg, type 2 NKT cells, and tumor associated macrophages (TAMs) (Najjar and Finke 2013). More recently, a group of cells named myeloidderived suppressor cells (MDSCs) has been considered as responsible for suppressing adaptive immunity and mediating pathological effects seen in MDS (Sica and Massarotti 2017; Chen et al. 2013; Kittang et al. 2015; Eksioglu et al. 2017; Sarhan et al. 2018).

2 MDSC: Pathways of Activation and Pathogenesis in MDS

Myeloid-derived suppressor cells (MDSCs) are a heterogeneous population of immune cells that are defined by their myeloid origin. This group of cells strongly expands in pathological situations such as chronic infections and solid tumors, suppressing immune mediated tumor surveillance and T cell polarization (Marvel and Gabrilovich 2015; Gabrilovich et al. 2007). MDSCs also share other non-immunological functions such as promotion of angiogenesis, tumor local invasion and metastases (Filipazzi et al. 2012; Chesney et al. 2017).

Despite the fact that MDSCs are a phenotypically heterogeneous population of myeloid cells at different stages of maturation, one feature common to all MDSCs is that they show remarkable activity to suppress various non-myeloid immune cells, such as T-cells, B-cells and natural killer (NK) cells (Filipazzi et al. 2012; Salminen et al. 2019a). MDSCs have also been reported to regulate innate immune responses by modulating the cytokine production of macrophages (Raza and Galili 2012; Lopez-Bujanda and Drake 2017). Three main subdivisions of MDSC have been proposed (Bronte et al. 2016): PMN-MDSCs or G-MDSCs (polymorphonuclear or granulocytic) which account for 70-80% of the MDSC population, M-MDSCs (monocytic) which account for 20-30% of MDSCs, and a smaller fraction of early-stage MDSC (eMDSC) (Lopez-Bujanda and Drake 2017; Bronte et al. 2016). There are some phenotypical differences when human and murine markers are compared. In humans, PMN-MDSCs are identified by the CD11b+CD14-CD15+ expression pattern or CD11b⁺CD14⁻CD66b⁺, while M-MDSC are characterized by CD11b+CD14+HLA^{low/-}CD15and eMDSC as lineage (CD3/14/15/19/56)-negative/HLA-DR⁻/CD33⁺ (Filipazzi et al. 2012; Bronte et al. 2016; Safarzadeh et al. 2019).

Physiologically, MDSCs are present predominantly in the bone marrow (Nadal et al. 2018). In pathological conditions, such as with cancer, chronic inflammation or autoimmunity, а sustained and aberrant differentiation of myeloid cells occurs, leading to MDSC expansion. Inflammatory factors can modulate myeloid cells in the tumor microenvironment, and having them delivered distantly to hematopoietic organs can change normal myelopoiesis and skew the differentiation of myeloid cells in favor of MDSCs (Umansky et al. 2016). The expansion and activation of MDSC are controlled by a complex network of soluble factors like IL-6, IL-10, IL-1 β and IFN- γ , granulocyte-macrophage colony stimulating factor (GM-CSF), M-CSF, G-CSF, vascular endothelial growth factor (VEGF) and TLR ligands (Gabrilovich and Nagaraj 2009; Veglia et al. 2018; Kumar et al. 2016a). Under pathological conditions and mediated by these factors, MDSC can be found in higher proportions in the blood circulation and may be also recruited back to the tumor microenvironment (Kumar et al. 2016b). Moreover, a tumor-microenvironment (TME) that is hypoxic, nutrient-deprived and enriched

in pro-inflammatory and suppressive cytokines, chemokines, and oxidative agents such as reactive oxygen species (ROS), nitric oxide (NO) and peroxynitrite, further induces the activation of local MDSCs (Tcyganov et al. 2018; Wang et al. 2013; Consonni et al. 2019; Weber et al. 2018).

MDSC are also defined by their functional ability to suppress immune cell responses (Bronte et al. 2016). This is achieved through the expression of many immune suppressive factors as, for instance, arginase (ARG1), NO and ROS. Unrestrained MDSC activation may, in turn cause DNA mutations and genetic instability.

The specific types of MDSCs use different mechanisms of immunosuppression. The essential difference is that the suppressive PMN-MDSCs produce ROS and ARG1, whereas M-MDSCs predominantly express the inducible nitric oxide synthase (iNOS). However, the common and main mechanism of action associated with the immunosuppressive activities of PMN-MDSCs and M-MDSCs is the metabolic conversion of L-arginine (L-Arg) through either iNOS or ARG1. Because both promote the depletion of the amino acid L-Arg and down-regulation of T cell receptor (TCR) ζ -chain expression, this leads to suppression of the cell cycle and T-cell immunosuppression as a result (Consonni et al. 2019).

MDSC expansion therefore inhibits proliferation and antitumor activity of T cells, decreasing cytokine secretion, recruiting regulatory T cells, and consequently, prohibiting natural killer cell (NK cell) activation, thus hampering the host anti-tumor immune response. Furthermore, MDSC also induce Treg differentiation by secreting IL-10 and TGF- β as well as stimulating tumor angiogenesis by secreting VEGF and basic fibroblast growth factor (Gabrilovich and Nagaraj 2009; Gabrilovich et al. 2012; Schmid and Varner 2012).

Clinical and experimental evidence has shown an association between MDSCs and cancer. A significant increase in MDSCs provokes a propitious immune microenvironment associated with high cancer prevalence, poor prognosis and resistance to therapy (Marvel and Gabrilovich 2015; Kirkwood et al. 2018; Lisha et al. 2018). On the other hand, the impact of age on MDSCs in humans is not well documented. The mechanisms involved in age-related increase of MDSCs seems to be, at least partly, determined by well-known aging-associated processes, including cellular senescence and chronic low-grade inflammation ("inflammaging"), and likely the skewing of hematopoiesis away from the lymphoid toward the myeloid lineage (Kirkwood et al. 2018; Pawelec et al. 2019; Salminen et al. 2019b). However, the recognition of expanded MDSC in MDS may offer a further genetic connection to aging (Chen et al. 2013).

In 2013, it was reported that MDSC were markedly expanded in the blood (Jiang et al. 2013) and bone marrow (Chen et al. 2013) of MDS patients. The latter study also demonstrated a suppressive effect of MDS MDSC on human erythroid and myeloid progenitor cell growth in vitro, implicating MDSC in the ineffective hematopoiesis associated with MDS (Chen et al. 2013). Moreover, with murine models, it was demonstrated that MDSC expansion is driven by pro-inflammatory S100A9, signalling through CD33 and mediated by the induction of immunosuppressive IL-10 and TGF- β (Chen et al. 2013). Independent studies subsequently revealed associations between MDSC and Treg expansion and higher MDS clinical risk (Kittang et al. 2015). Novel therapies are now under consideration, directed towards MDSC and with the goal of ameliorating immunosuppression and improving hematopoiesis (Eksioglu et al. 2017; Sarhan et al. 2018). However, the relationship between these immune cell populations, inflammatory signaling and recurring mutations in MDS is not completely understood. The influence of different factors in expansion and activation of MDSCs such as PGE2, VEGF, IL-6, IL-10 and S100A8-A9 has been shown (Rosenberg and Sinha 2009; Marvel and Gabrilovich 2015; Najjar and Finke 2013; Gabrilovich and Nagaraj 2009; Pawelec et al. 2019). Due to the profile of inflammatory cytokine changes with the course of disease, current studies have focused on understanding the abnormalities in immunologic profile in different myeloid disorders, mainly in MDS and AML, and other proposals to deplete the key innate immune cellular effectors, MDSC, are still currently in development (Sallman and List 2019; Pawelec et al. 2019).

3 Considerations for Clonal, Pre-MDS States: CHIP and CCUS

Major breakthroughs in the molecular and genetic basis of MDS have recently been achieved. Genetic lesions, namely recurrent somatic point mutations and/or small insertions/deletions in >40 different genes, have now been associated with MDS pathogenesis (Cazzola 2020; Cull and Rauh 2017; Claus and Lubbert 2003; Issa 2010; Sperling et al. 2017). These mainly occur in genes regulating RNA splicing (e.g., SF3B1, SRSF2, U2AF1, ZRSR2), DNA methylation (e.g., TET2, DNMT3A, IDH1/IDH2) and chromatin accessibility (e.g., ASXL1, EZH2, STAG2, RAD21), transcription factors (e.g. RUNX1, GATA2, ETV6), signal transduction (e.g. CBL, JAK2, NRAS) and the response to DNA damage (e.g. TP53, PPM1D). In the case of SF3B1, the presence of mutations is incorporated into the current iteration of the World Health Organization classification of MDS, with a recent proposal to recognized SF3B1-mutant MDS as an even more distinct diagnostic entity (Malcovati et al. 2020).

Presently, other MDS-associated mutations are not considered diagnostic of MDS on their own. This is because at least 10–15% of healthy older persons with no hematologic disease acquire somatic mutations that overlap with MDS, drive clonal expansion and, eventually, what is now called clonal hematopoiesis of indeterminate potential (CHIP) (Steensma et al. 2015). Although most individuals who acquire CHIP during aging will never develop MDS, the presence of an MDS-associated somatic mutation, such as in DNMT3A, ASXL1 or TP53 is a predictor of the development strong of subsequent hematologic malignancy and is associated with worse overall survival (Park et al. 2019; Yoshizato et al. 2015). The presence of а recurrent mutation and otherwise



Fig. 1 Some of the main pathways involved in MDS pathogenesis and progression. An aging and inflammatory bone marrow microenvironment induced by cellular senescence and chronic immune stimulation leads to MDSC accumulation and activation. Inflammatory cytokines and soluble factors, such as ROS and iNOS, contribute to increased apoptosis and pyroptosis of HSPCs as well as genomic instability. Somatic mutations contribute to further dysregulation of immune system,



suppression of normal hematopoiesis, clonal evolution and susceptibility to leukemic transformation. *HSPC* Hematopoietic stem and progenitor cells, *IMCs* Immature myeloid cells; *MDSCs* Myeloid-derived suppressor cells, *ROS* Reactive oxygen species, *iNOS* Inducible nitric oxide synthase, *TNF-a* Tumor necrosis factor-alpha, *IFN-γ* Interferon gamma, *TET2* Tet methylcytosine dioxygenase 2, *DNMT3A* DNA methyltransferase 3A, *ASXL1* Additional sex combs like 1, *JAK2* Janus Kinase 2

unexplained cytopenias (clonal cytopenias of undetermined significance, CCUS) can be suggestive of progression to MDS, although in this case more specific morphologic criteria are mandatory (Sperling et al. 2017; Cargo et al. 2015; Kwok et al. 2015). A subsequent study with longitudinal follow-up suggests it may be possible to predict patients with CCUS at greatest risk of progression to MDS or other myeloid neoplasm and that, effectively, CCUS patients with particular mutational features may have presumptive evidence of early MDS (Malcovati et al. 2017).

As described, there is a complex process involved in the pathogenesis of MDS, especially mechanisms directly correlated with myeloidmediated inflammation, the accumulation of genetic damage, immunosuppression and related selective pressures during the evolution of malignant clones (Steensma et al. 2015; Hosono 2019) (Fig. 1). Although we have mainly focused on these aspects in the context of MDS, these processes are likely at play as early as the CHIP phase and into CCUS. For example, common drivers of CHIP and MDS, mutations in TET2 and DNMT3A, have been associated with inflammation and immune cell alterations in humans and murine models, as recently reviewed (King et al. 2020; SanMiguel et al. 2020; Ferrone et al. 2020; Cook et al. 2020). Moreover, spliceosomal

mutations (e.g. SF3B1, SRSF2, U2AF1) also appear to converge on innate immune pathways, resulting in excessive inflammasome activation and inflammatory cytokine production, including IL-6 (Smith et al. 2019; Pollyea et al. 2019). Finally, our group has demonstrated increased ARG1 expression in Tet2-mutant macrophages (Cull et al. 2017) and in MDS patients with DNMT3A and TET2 mutations (Cull et al. 2018). This could signify a myeloid suppressive phenotype (reminiscent of MDSCs) in response to chronic inflammation but more studies are required to map the relationship between immune dysregulation and mutations in the progression from CHIP to CCUS and MDS. The comprehension of genetic lesions, genomic instability and dysregulated immune response are important to better understand MDS pathogenesis, progression and predictive factors of response to therapy.

4 Conclusions

Though the pathogenesis in MDS is now much more understood, the factors involved in this heterogeneous process are still an attractive and constant focus of research. Clinical and experimental evidence suggest an important link between genetic, epigenetic, and immune systems in the pathogenesis and progression of MDS. Ongoing studies are looking at more specific molecular and immune pathways and targets with potential clinical significance, notably the role of the inflammatory marrow environment and MDSCs in MDS. Translation from understanding the complex molecular and immunological pathophysiology of MDS to the identification of new targets and novel treatment options are long awaited.

References

- Banerjee T, Calvi LM, Becker MW et al (2019) Flaming and fanning: the spectrum of inflammatory influences in myelodysplastic syndromes. Blood Rev 36:57–69
- Barreyro L, Chlon TM, Starczynowski DT (2018) Chronic immune response dysregulation in MDS pathogenesis. *Blood* 132(15):1553–1560
- Basiorka AA, McGraw KL, Eksioglu EA, Chen X, Johnson J, Zhang L, Zhang Q, Irvine BA, Cluzeau T, Sallman DA, Padron E, Komrokji R, Sokol L, Coll RC, Robertson AA, Cooper MA, Cleveland JL, O'Neill LA, Wei S, List AF (2016) The NLRP3 inflammasome functions as a driver of the myelodysplastic syndrome phenotype. Blood 128(25):2960–2975
- Bronte V, Brandau S, Chen SH et al (2016) Recommendations for myeloid-derived suppressor cell nomenclature and characterization standards. Nature 7:12150
- Cargo CA, Rowbotham N, Evans PA et al (2015) Targeted sequencing identifies patients with preclinical MDS at high risk of disease progression. Blood 126 (21):2362–2365
- Cazzola M (2020) Myelodysplastic syndromes. N Engl J Med 383(14):1358–1374
- Chen X, Eksioglu EA, Zhou J, Zhang L, Djeu J, Fortenbery N, Epling-Burnette P, Van Bijen S, Dolstra H, Cannon J, Youn J, Donatelli SS, Qin D, De Witte T, Tao J, Wang H, Cheng P, Gabrilovich DI, List A, Wei S (2013) Induction of myelodysplasia by myeloid-derived suppressor cells. J Clin Invest 123 (11):4596–4611
- Chesney JA, Mitchell RA, Yaddanapudi K (2017) Myeloid-derived suppressor cells- a new therapeutic target to overcome resistance to cancer immunotherapy. J Leukoc Biol 102(3):727–740
- Claus R, Lubbert M (2003) Epigenetic targets in hematopoietic malignancies. Oncogene 22 (42):6489–6496
- Consonni FM, Porta C, Marino A et al (2019) Myeloidderived suppressor cells: ductile targets in disease. Front Immunol 10:949
- Cook EK, Luo M, Rauh MJ (2020) Clonal hematopoiesis and inflammation: partners in leukemogenesis and comorbidity. Exp Hematol 83:85–94

- Corey SJ, Minden MD, Barber DL et al (2007) Myelodysplastic syndromes: the complexity of stemcell diseases. Nat Rev Cancer 7(2):118–129
- Cull AH, Rauh MJ (2017) Success in bone marrow failure? Novel therapeutic directions based on the immune environment of myelodysplastic syndromes. J Leukoc Biol 102(2):209–219
- Cull AH, Snetsinger B, Buckstein R, Wells RA, Rauh MJ (2017) Tet2 restrains inflammatory gene expression in macrophages. Exp Hematol 55:56–70.e13
- Cull AH, Mahendru D, Snetsinger B et al (2018) Overexpression of Arginase 1 is linked to DNMT3A and TET2 mutations in lower-grade myelodysplastic syndromes and chronic myelomonocytic leukemia. Leuk Res 65:5–13
- de Matos AG, Ribeiro Junior HL, de Paula BD et al (2017) Interleukin-8 and nuclear factor kappa B are increased and positively correlated in myelodysplastic syndrome. Med Oncol 34(10):168
- Eksioglu EA, Chen X, Heider K-H, Rueter B, McGraw KL, Basiorka AA, Wei M, Burnette A, Cheng P, Lancet J, Komrokji R, Djeu J, List A, Wei S (2017) Novel therapeutic approach to improve hematopoiesis in low risk MDS by targeting MDSCs with the Fc-engineered CD33 antibody BI 836858. Leukemia 31(10):2172–2180
- Ferrone CK, Blydt-Hansen M, Rauh MJ (2020) Age-associated TET2 mutations: common drivers of myeloid dysfunction, cancer and cardiovascular disease. Int J Mol Sci 21(2):626
- Filipazzi P, Huber V, Rivoltini L (2012) Phenotype, function and clinical implications of myeloid-derived suppressor cells in cancer patients. Cancer Immunol Immunother 61(2):255–263
- Gabrilovich DI, Nagaraj S (2009) Myeloid-derived-suppressor cells as regulators of the immune system. Nat Rev Immunol 9(3):162–174
- Gabrilovich DI, Bronte V, Chen SH et al (2007) The terminology issue for myeloid-derived suppressor cells. Cancer Res 67(1):425–426
- Gabrilovich DI, Ostrand-Rosenberg S, Bronte V (2012) Coordinated regulation of myeloid cells by tumours. Nat Rev Immunol 12(4):253–268
- Ganan-Gomez I, Wei Y, Starczynowski DT et al (2015) Deregulation of innate immune and inflammatory signaling in myelodysplastic syndromes. Leukemia 29 (7):1458–1469
- Glenthoj A, Orskov AD, Hansen JW et al (2016) Immune mechanisms in myelodysplastic syndrome. Int J Mol Sci 17(6):944
- Hosono N (2019) Genetic abnormalities and pathophysiology of MDS. Int J Clin Oncol 24(8):885–892
- Issa JP (2010) Epigenetic changes in the myelodysplastic syndrome. Hematol Oncol Clin North Am 24 (2):317–330
- Jiang HJ, Fu R, Wang HQ, Li LJ, Qu W, Liang Y, Wang GJ, Wang XM, Wu YH, Liu H, Song J, Guan J, Xing LM, Ruan EB, Shao ZH (2013) Increased circulating of myeloid-derived suppressor cells in myelodysplastic syndrome. Chin Med J 126(13):2582–2584

- Kawai T, Akira S (2010) The role of pattern-recognition receptors in innate immunity: update on toll-like receptors. Nat Immunol 11(5):373–384
- Kerbauy DB, Joachim H (2007) Apoptosis and antiapoptotic mechanisms in the progression of MDS. Exp Hematol 35(11):1739–1746
- King KY, Huang Y, Nakada D, Goodell MA (2020) Environmental influences on clonal hematopoiesis. Exp Hematol 83:66–73
- Kirkwood KL, Zhang L, Thiyagarajan R et al (2018) Myeloid-derived suppressor cells at the intersection of inflammaging and bone fragility. Immunol Investig 47:844–854
- Kittang AO, Kordasti S, Sand KE, Constantini B, Kramer AM, Perezabellan P, Seidl T, Rye KP, Hagen KM, Kulasekararaj A, Bruserud O, Mufti GJ (2015) Expansion of myeloid-derived suppressor cells correlates with number of T regulatory cells and disease progression in myelodysplastic syndrome. Onco Targets Ther 5(2):e1062208
- Kornblau SM, McCue D, Singh N et al (2010) Recurrent expression signatures of cytokines and chemokines are present and are independently prognostic in acute myelogenous leukemia and myelodysplasia. Blood 116 (20):4251–4261
- Kumar V, Patel S, Tcyganov E, Gabrilovich D (2016a) The nature of myeloid-derived suppressor cells in the tumor microenvironment. Trends Immunol 37 (3):208–220
- Kumar V, Patel S, Toyganov E, Gabrilovich DI (2016b) The nature of myeloid-derived suppressor cells in the tumor microenvironment. Trends Immunol 37 (3):208–220
- Kwok B, Hall JM, Witte S et al (2015) MDS-associated somatic mutations and clonal hematopoiesis are common in idiopathic cytopenias of undetermined significance. Blood 126(21):2355–2361
- Lisha A, Mu S, Wang Y et al (2018) Prognostic role of myeloid-derived suppressor cells in cancers: a systematic review and meta-analysis. BMC Cancer 18:1220
- Lopez-Bujanda Z, Drake CG (2017) Myeloid-derived cells in prostate cancer progression: phenotype and prospective therapies. J Leukoc Bio 102(2):393–406
- Malcovati L, Galli A, Travaglino E, Ambaglio I, Rizzo E, Molteni E, Elena C, Ferretti VV, Catricala S, Bono E, Todisco G, Bianchessi A, Rumi E, Zibellini S, Pietra D, Boveri E, Camaschella C, Toniolo D, Papaemmanuil E, Ogawa S, Cazzola M (2017) Clinical significance of somatic mutation in unexplained blood cytopenia. Blood 129(25):3371–3378
- Malcovati L, Stevenson K, Papaemmanuil E, Neuberg D, Bejar R, Boultwood J, Bowen DT, Campbell PJ, Ebert BL, Fenaux P, Haferlach T, Heuser M, Jansen JH, Komrokji RS, Maciejewski JP, Walter MJ, Fontenay M, Garcia-Manero G, Graubert TA, Karsan A, Meggendorfer M, Pellagatti A, Sallman DA, Savona MR, Sekeres MA, Steensma DP, Tauro S, Thol F, Vyas P, Van de Loosdrecht AA, Haase D, Tuchler H, Greenberg PL, Ogawa S,

Hellstrom-Lindberg E, Cazzola M (2020) SF3B1mutant MDS as a distinct disease subtype: a proposal from the International Working Group for the Prognosis of MDS. Blood 136(2):157–170

- Marvel D, Gabrilovich DI (2015) Myeloid-derived suppressor cells in the tumor microenvironment: expect the unexpected. J Clin Invest 125(9):3356–3364
- Muto T, Walker CS, Choi K, Hueneman K, Smith MA, Gul Z, Garcia-Manero G, Ma A, Zheng Y, Starczynowski DT (2020) Adaptive response to inflammation contributes to sustained myelopoiesis and confers a competitive advantage in myelodysplastic syndrome HSCs. Nat Immunol 21 (5):535–545
- Nadal C, Beguin J, Benchekroun G, Le Roux D (2018) The myeloid derived suppressor cells: Who are they? Can they be used as a diagnostic tool to investigate metastasis in veterinary medicine? Comp Immunol Microbiol Infect Dis 61:5–8
- Najjar YG, Finke JH (2013) Clinical perspectives on targeting of myeloid derived suppressor cells in the treatment of cancer. Front Oncol 3:49
- Park DS, Akuffo AA, Muench DE et al (2019) Clonal hematopoiesis of indeterminate potential and its impact on patient trajectories after stem cell transplantation. PLoS Comput Biol 15:e1006913
- Pawelec G, Verschoor CP, Ostrand-Rosenberg S (2019) Myeloid-derived suppressor cells: not only in tumor immunity. Front Immunol 10:1099
- Pollyea DA, Harris C, Rabe JL, Hedin BR, De Arras L, Katz S, Wheeler E, Bejar R, Walter MJ, Jordan CT, Pietras EM, Alper S (2019) Myelodysplastic syndrome-associated spliceosome gene mutations enhance innate immune signaling. Haematologica 104(9):e388–e392
- Raza A, Galili N (2012) The genetic basis of phenotypic heterogeneity in myelodysplastic syndromes. Nat Rev Cancer 12(12):849–859
- Rosenberg SO, Sinha P (2009) Myeloid-derived suppressor cells: linking inflammation and cancer. J Immunol 182(8):4499–4506
- Safarzadeh E, Hashemzadeh S, Duijf PHG et al (2019) Circulating myeloid-derived suppressor cells: an independent prognostic factor in patients with breast cancer. J Cell Physiol 234(4):3515–3525
- Sallman DA, List A (2019) The central role of inflammatory signaling in the pathogenesis of myelodysplastic syndromes. Blood 133(10):1039–1048
- Salminen A, Kaarniranta K, Kauppinen A (2019a) Immunosenescence: the potential role of myeloidderived suppressor cells (MDSC) in age-related immune deficiency. Cell Mol Life Sci 76 (10):1901–1918
- Salminen A, Kauppinen A, Kaarniranta K (2019b) AMPK activation inhibits the functions of myeloid-derived suppressor cells (MDSC): impact on cancer and aging. J Mol Med 97(8):1049–1064
- SanMiguel JM, Young K, Trowbridge JJ (2020) Hand in hand: intrinsic and extrinsic drivers of aging and clonal

hematopoiesis. Exp Hematol:S0301-472X(20) 30562–2

- Sarhan D, Brandt L, Felices M, Guldevall K, Lenvik T, Hinderlie P, Curtsinger J, Warlick E, Spellman SR, Blazar BR, Weisdorf DJ, Cooley S, Vallera DA, Onfelt B, Miller JS (2018) 161533 TriKE stimulates NK-cell function to overcome myeloid-derived suppressor cells in MDS. Blood Adv 2(12):1459–1469
- Schmid MC, Varner JA (2012) Myeloid cells in tumor inflammation. Vasc Cell 4(1):14
- Sekeres MA (2010) The epidemiology of myelodysplastic syndromes. Hematol Oncol Clin North Am 24 (2):287–294
- Xin S, Yanhua Z, Li X et al (2019) The inflammatory cytokine profile of myelodysplastic syndromes. A meta-analysis. Medicine 98:22 (e15844
- Sica A, Massarotti M (2017) Myeloid suppressor cells in cancer and autoimmunity. J Autoimmun 85:117–125
- Sloand EM, Melenhorst JJ, Tucker ZC, Pfannes L, Brenchley JM, Yong A, Visconte V, Wu C, Gostick E, Scheinberg P, Olnes MJ, Douek DC, Price DA, Barrett AJ, Young NS (2011) T-cell immune responses to Wilms tumor 1 protein in myelodysplasia responsive to immunosuppressive therapy. Blood 117 (9):2691–2699
- Smith MA, Choudhary GS, Pellagatti A, Choi K, Bolanos LC, Bhagat TD, Gordon-Mitchell S, Von Ahrens D, Pradhan K, Steeples V, Kim S, Steidl U, Walter M, Fraser IDC, Kulkarni A, Salomonis N, Komurov K, Boultwood J, Verma A, Starczynowski DT (2019) U2AF1 mutations induce oncogenic IRAK4 isoforms and activate innate immune pathways in myeloid malignancies. Nat Cell Biol 21(5):640–650
- Sperling AS, Gibson CJ, Ebert BL (2017) The genetics of myelodysplastic syndrome: from clonal hematopoiesis to secondary leukemia. Nat Rev Cancer 17:5–19
- Starczynowski DT, Kuchenbauer F, Argiropoulos B, Sung S, Morin R, Muranyi A, Hirst M, Hogge D, Marra M, Wells RA, Buckstein R, Lam W, Humphries RK, Karsan A (2010) Identification of miR-145 and miR-146a as mediators of the 5q- syndrome phenotype. Nat Med 16(1):49–58
- Steensma DP, Bejar R, Jaiswal S, Lindsley RC, Sekeres MA, Hasserjian RP et al (2015) Clonal hematopoiesis

of indeterminate potential and its distinction from myelodysplastic syndromes. Blood 126(1):9-16

- Tcyganov E, Mastio J, Chen E, Gabrilovich DI (2018) Plasticity of myeloid-derived suppressor cells in cancer. Curr Opin Immunol 51:76–82
- Umansky V, Blattner C, Gebhardt C et al (2016) The role of myeloid-derived suppressor cells (MDSC) in cancer progression. Vaccines (Basel) 4(4):pii: E36
- Valka J, Veselaa J, Votavova H et al (2019) Genetic variant screening of DNA repair genes in myelodysplastic syndrome identifies a novel mutation in the XRCC2 gene. Oncol Res Treat 42:263–268
- Varney ME, Niederkorn M, Konno H, Matsumura T, Gohda J, Yoshida N, Akiyama T, Christie S, Fang J, Miller D, Jerez A, Karsan A, Maciejewski JP, Meetei RA, Inoue J, Starczynowski DT (2015) Loss of Tifab, a del(5q) MDS gene, alters hematopoiesis through derepression of toll-like receptor-TRAF6 signaling. J Exp Med 212(11):1967–1985
- Veglia F, Perego M, Gabrilovich D (2018) Myeloidderived suppressor cells coming of age. Nat Immunol 19(2):108–119
- Wang L, Chang WY, Wong SC et al (2013) Increased myeloid-derived suppressor cells in gastric Cancer correlate with Cancer stage and plasma S100A8/A9 Proinflammatory proteins. J Immunol 190(2):794–804
- Wang C, Yang Y, Gao S et al (2018) Immune dysregulation in myelodysplastic syndrome: clinical features, pathogenesis and therapeutic strategies. Crit Rev Oncol Hematol 122:123–132
- Weber R, Fleming V, Hu X et al (2018) Myeloid-derived suppressor cells hinder the anti-cancer activity of immune checkpoint inhibitors. Front Immunol 9:1310
- Winter S, Shoaie S, Kordasti S, Platzbecker U (2020) Integrating the "Immunome" in the stratification of myelodysplastic syndromes and future clinical trial design. J Clin Oncol 38(15):1723–1735
- Yang L, Qian Y, Eksioglu E et al (2015) The inflammatory microenvironment in MDS. Cell Mol Life Sci 72 (10):1959–1966
- Yoshizato T, Dumitriu D, Hosokawa K et al (2015) Somatic mutations and clonal hematopoiesis in aplastic anemia. N Engl J Med 373:35–47



Stem Cell Aging and Regenerative Medicine

Debojyoti De, Parimal Karmakar, and Debalina Bhattacharya

Abstract

Stem cells are a promising source for regenerative medicine to cure a plethora of diseases that are currently treated based on either palliative or symptomatic relief or by preventing their onset and progression. Aging-associated degenerative changes in stem cells, stem cell niches, and signaling pathways bring a step by step decline in the regenerative and functional potential of tissues. Clinical studies and experiments on model organisms have pointed out checkpoints that aging will inevitably impose on stem cell aiming for transplantation and hence questions are raised about the age of the donor. In the following discourse, we review the fundamental molecular pathways that are implicated in stem cell aging and the current progress in tissue engineering and transplantation of each type of stem cells in regenerative medicine. We further focus on the consequences of stem cell aging on their clinical uses and the development of novel strategies to bypass those pitfalls and improve tissue replenishment.

D. Bhattacharya (🖂)

Keywords

 $\begin{array}{l} Aging \cdot Regeneration \cdot Regenerative \\ medicine \cdot Senescence \cdot Stem cells \end{array}$

Abbreviations

ADSC	Adipose tissue-derived stem cell			
ASC	Adult stem cell			
BMSC/	Bone marrow-derived mesenchy-			
BMDSC	mal stem/stromal cell			
CSC	Cardiac stem cell			
DNMT	DNA methyltransferase			
EPC	Endothelial progenitor cell			
ESC	Embryonic stem cell			
G-CSF	Granulocyte colony-stimulating			
	factor			
GM-CSF	Granulocyte-macrophage colony-			
	stimulating factor			
GSC	Germline stem cell			
GVHD	Graft versus host disease			
HAT	Histone acetyltransferase			
HDAC	Histone deacetylase			
HIF	Hypoxia-Inducible Factor			
HSC	Haematopoietic stem cell			
ICM	Inner cell mass			
iPSC	Induced pluripotent stem cell			
ISC	Intestinal stem cell			
LDHA	Lactate dehydrogenase A			
MDSC	Muscle-derived stem cell			
MSC	Mesenchymal stem cell			
NAC	N-acetylcysteine			

D. De and P. Karmakar

Department of Life science and Biotechnology, Jadavpur University, Kolkata, India

Department of Microbiology, Maulana Azad College, Kolkata, India

e-mail: debalina.bhattacharya13@gmail.com; debalina.bhattacharya@maulanaazadcollegekolkata.ac.in